

ANDREW GORBA:



EXHIBIT

1

BBMM +

Received 40 cc cold blood (w/ CPD)

PE monos 30%
Poly 70%

IL3 SANDO
Went

IL6 SANDO
Went

SCF ANGEL
Went

added 15 ml 1xPBS to bring to incubation volume of 75 ml.

Added 1.5 ml antibody (12.8)
incubated 25 ml.

Primed CellPro "ceprate".

final concn
will be dil
Cells are
concentrated

spin cells. Resuspended in 1x PBS to a final volume of 300 cc in bag.

Ran through column.

BBMM: FB
BS

Unadsorbed portion → spun down.
and consolidated in 1x PBS
for incubation.

2.8×10^6 cells
Put in

75 ml for incubation (added HPA)
1.5 ml antibody (12.8) 25 min. Spun down
following incubation. Resp to vol. of 300 cc in bag
Ran through 2nd column.

stem cell portions from Runs 1 & 2
were combined (after counts done and
samples removed for staining)

total cells 2.8×10^6 for transduction

BBMM + 31615cf (for 500ml of media)

1L3 SANDOZ * 10230092 stock at 150ug/ml
want final: 20ug/ml x 2 ... 20ug

133ul add

1L6 SANDOZ * 10150392 stock at 150ug/ml
want final: 50ug/ml x 2 ... 50ug

333ul add

SLF AUBIEN * 1509F2 stock at 1.5mg/ml = 1500ug/ml
want final: 100ug/ml x 2 ... 100ug

167ul add

final concentrations are doubled since the media
will be diluted 1:2 w/ viral supernatant.
Cells are therefore incubated with the correct
concentrations.

BBMM: FBS Gemini lot# A2003H
BSA #115

2.8×10^6 cells want final: 5×10^4 cells.

Put in 2 T75 30ml each: 15ml B365 051193

15ml LASU^{G7} lot# 53

+ protamine sulfate 240ul
of 1:10 diluted 50

in down
300ul in bag.

42
2ml

Cord Blood cells pre processing:

CFUs

SET 143

Start:

Plate #	Sample	# Cells	# ul/ml media
-G418	1ab	5×10^4	50
+G418	2ab		50
-G418	3ab	1×10^5	100
+G418	4ab		100

adsorb
fraction

adsorb
fraction

CFUs Post transduction: SET 144

plate #	# Cells	# cell
-G418	500	7
↓	1000	14
+G418	2000	28
↓	500	7
	1000	14
	2000	28

(yields)
adsorb

count:

$$\bar{x} = 34$$

$$\times 2 \times 10^4 = 6.8 \times 10^5 \text{ Clm1}$$

$$\times 5.5 \text{ ml} = 3.7 \times 10^6 \text{ C}$$

adsorb
fraction from
media

Reinfused on 5/15/93
No transduced stem cells

G418

-

+

7ab
8ab

1000
2000

20
40

Start:

$5 \times 10^8 \text{ c}$

PRE
 0.71%

Post ab
 0.22%

*ul/ml media

$*34+ = 3.6 \times 10^6 \text{ c} = 1.1 \times 10^6 \text{ c}$

adsorbed
fraction #1:

$2 \times 10^6 \text{ c}$

F1 F2 gate
 31.94%

F1 F2 gate
 20.81%

$*34+ = 0.64 \times 10^6 \text{ c} = 0.42 \times 10^6 \text{ c}$

adsorbed
fraction #2:

$0.8 \times 10^6 \text{ c}$

2.46%

5.80%

$\pm 34+c = 0.02 \times 10^6 \text{ c}$

$0.05 \times 10^6 \text{ c}$

(yields)

adsorbed #1:

$$\frac{\text{PRE} \& \text{F1/F2 gate}}{0.64 \times 10^6 \text{ c}} = \boxed{17.8\%}$$

$$\frac{\text{PRE} \& \text{F1/F2 gate}}{0.42 \times 10^6 \text{ c}} = \boxed{11.7\%}$$

$$\frac{\text{Post ab} \& \text{F1/F2 gate}}{0.64 \times 10^6 \text{ c}} = \boxed{58.2\%}$$

$$\frac{\text{Post ab} \& \text{F1/F2 gate}}{0.42 \times 10^6 \text{ c}} = \boxed{38.2\%}$$

adsorbed #2:

PRE & F1/F2

PRE & F1/F2

~~removed from~~
~~medium~~~~medium~~

ZACHARY RIGGINS:

5/14/93

REC'D 200cc COLD BLOOD

PRE: $\frac{\text{mon}}{109}$ \checkmark $\frac{\text{poly}}{109}$

$$218 \times 50 = 10.9 \times 10^6 \text{ clml}$$

$$\times 200 \text{ ml} = 2.2 \times 10^9 \text{ C} \quad \text{start}$$

Added 3 vials (4.5 ml) 12.8 ab.
inc. 25 min.

Spindown. Rsp'd. in 1x PBS to 300ml
in bag.

Ran through column:

spin down unadsorbed fraction for 2nd ab
incubation.

Spin stem cell fraction to Rsp'd in
smaller volume for count.

Counts:

unadsorbed

mon \checkmark poly
67 102

$$10.9 \times 50 \times 10^3$$

$$= 8.5 \times 10^6 \text{ clml} \times 225 \text{ ml}$$

$$= 1.9 \times 10^9 \text{ C}$$

stem

mon \checkmark poly
172 16

$$188 \times 2 \times 10^4$$

$$= 3.8 \times 10^6 \text{ clml} \times 5.5 \text{ ml}$$

$$= 20.7 \times 10^6 \text{ C}$$

incl
12.8
SPU
Pot
Ran

Cor

1

2

3

3.

=

Per
free

com

26x1

was

= 2 =

13 fl

LA 51

incubated unadsorbed fraction w/ 4.5 ml
 12.8 ab. for 25 min.
 spun down.

Put in 300ml in bag (w/ 1x PBS)
 Ran through 2nd column.

counts:

unadsorbed
 monos polys
 30 33

$$63 \times 53 \times 10^3$$

$$3.15 \times 10^6 \text{ cpm} \times 600 \text{ ml}$$

$$= 1.9 \times 10^9 \text{ c}$$



stem
 monos polys
 58 4

$$62 \times 2 \times 10^4$$

$$= 1.2 \times 10^6 \text{ cpm}$$

$$\times 5 \text{ ml} = 6 \times 10^6 \text{ c}$$

percolated/ficoll'd
 freeze \Rightarrow LWT(2)

combined stem cell fractions

$2 \times 10^6 \text{ c}$ for transduction

want final $[T] = 5 \times 10^4 \text{ c/ml}$

520 ml total

$- 2 = 260 \text{ ml sup}$

260 ml media

13 flasks 40 ml/flask

20 ml sup

20 ml media (B36S)

+ 300 ml protamine sulfate

(2)

LAWS sup 539 (bottles 18/19)

$\times 10^4$
 ml $\times 55 \text{ ml}$

CFUs:

5/17/93

PRE

Plate #	Sample	# cells	# ml
lab	(-G418)	5×10^4	5
2ab	PRE trans ↓ (+G418)	↓	5

BBMM + 31615CF:

1L3 Sander #y0230392

1L6 Sander #y0450392

SCF AMGEN #150952

Took sample to micro for sterility ✓
 each day of transduction pt.
 Stat Gram stain done (negative)
 before cells were given to baby.

5/15/93 4pm 2nd transduction.
 Spun cells down from each flask
 Respt in fresh media & LASN sup
 added Protamine sulfate

5/16/93 3rd transduction 330pm.
 Repeated above.

5/17

COR

PRE

REX

POST

OG

+G4

OG

+G4

517193 cells washed 4x
 3x in 1x PBS + P15
 last wash in RPMI (no p15)

count: 60×10^6 c

$$\bar{x} = 15 \times 10 \times 10^4 = 15 \times 10^9$$

$$\times 40 \text{ ml} = 60 \times 10^6 \text{ c}$$

Put in 5cc into 10cc syringe

Reinfused on 517193 (LCSF)

Post trans. CPUs: 50746

Sample plate #		# cells	# ul
DG418 ↓ +G418 ↓ DG418 +G418	1ab	500	4
	2ab	1000	8
	3ab	2000	16
	4ab	500	4
	5ab	1000	8
↓ DG418 +G418	6ab	2000	16
	7ab	1000	24
	8ab	1000	24

ry ✓
 (wire)
 baby
 lion
 in flock
 SN supe

30pm